

01/02/98

10541 U.S. PTO

## HAMILTON, BROOK, SMITH &amp; REYNOLDS, P.C.

UTILITY  
PATENT APPLICATION  
TRANSMITTAL

(Only for new nonprovisional applications under 37 CFR 1.53(b))

Attorney Docket No.	GSF97-06	Total Pages	52
First Named Inventor or Application Identifier			
Gerd Sutter			
Express Mail Label No.	EM361199084US		

## APPLICATION ELEMENTS

See MPEP chapter 600 concerning utility patent application contents.

1. ☒ Fee Transmittal Form  
(Submit an original, and a duplicate for fee processing)
2. ☒ Specification [Total Pages 44]  
(preferred arrangement set forth below)
  - Descriptive title of the invention
  - Cross References to Related Applications
  - Statement Regarding Fed sponsored R & D
  - Reference to Microfiche Appendix
  - Background of the invention
  - Brief Summary of the invention
  - Brief Description of the Drawings (if filed)
  - Detailed Description
  - Claim(s)
  - Abstract of the Disclosure
3. ☐ 6 Drawing(s) (35 USC 113) [Total Sheets 6]
4. Oath or Declaration / POA [Total Pages]
  - a. ☐ Newly executed (original or copy)
  - b. ☐ Copy from a prior application (37 CFR 1.63(d))  
(for continuation/divisional with Box 17 completed)  
[Note Box 5 below]
  - c. ☐ DELETION OF INVENTOR(S)  
Signed statement attached deleting inventor(s) named in the prior application, see 37 CFR 1.63(d)(2) and 1.33(b).
5. ☐ Incorporation By Reference (useable if Box 4b is checked)  
The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.

ADDRESS TO: Assistant Commissioner for Patents  
Box Patent Application  
Washington, DC 20231

6. ☐ Microfiche Computer Program (Appendix)
7. Nucleotide and/or Amino Acid Sequence Submission  
(if applicable, all necessary)
  - a. ☐ Computer Readable Copy
  - b. ☐ Paper Copy (identical to computer copy)
  - c. ☐ Statement verifying identity of above copies

## ACCOMPANYING APPLICATION PARTS

8. ☐ Assignment Papers (cover sheet & document(s))
9. ☐ 37 CFR 3.73(b) Statement [ ] Power of Attorney  
(when there is an assignee)
10. ☐ English Translation Document (if applicable)
11. ☐ Information Disclosure Statement (IDS)/PTO-1449 [ ] Copies of IDS Citations
12. ☐ Preliminary Amendment
13. ☒ Return Receipt Postcard (MPEP 503)  
(Should be specifically itemized)
14. ☐ Small Entity [ ] Statement filed in prior application,  
Statement(s) [ ] Status still proper and desired
15. ☐ Certified Copy of Priority Document(s)  
(if foreign priority is claimed)
16. ☐ Other: .....

17. If a CONTINUING APPLICATION, check appropriate box and supply the requisite information:

☒ Continuation ☐ Divisional ☐ Continuation-in-part (CIP) of prior application No: PCT / EP96/02926

## 18. CORRESPONDENCE ADDRESS

☐ Customer Number or Bar Code Labelor ☒ Correspondence address below

(Insert Customer No. or Attach bar code label here)

NAME	David E. Brook				
	HAMILTON, BROOK, SMITH & REYNOLDS, P.C.				
ADDRESS	Two Militia Drive				
CITY	Lexington	STATE	MA	ZIP CODE	02173
COUNTRY	USA	TELEPHONE	(781)861-6240	FAX	(781)861-9540

SIGNATURE

Anne J. Collins

DATE

January 2, 1998

# HAMILTON, BROOK, SMITH & REYNOLDS, P.C.

<b>FEE TRANSMITTAL FOR PATENT APPLICATIONS</b>		Attorney Docket Number		GSF97-06	
		Application Number			
		First Named Inventor		Gerd Sutter	
CLAIM CALCULATION (includes any preliminary amendment)					
CLAIMS	(1) FOR	(2) NUMBER FILED	(3) NUMBER EXTRA	(4) RATE	(5) CALCULATIONS
	TOTAL CLAIMS <small>(37 CFR 1.16(c))</small>	30 - 20 =	10	x \$ 22 =	\$ 220.00
	INDEPENDENT CLAIMS <small>(37 CFR 1.16(b))</small>	3 - 3 =	0	x \$ 82 =	\$ 0
	MULTIPLE DEPENDENT CLAIMS (if applicable) <small>(37 CFR 1.16(d))</small>			+ \$ 270 =	\$
				BASIC FEE <small>(37 CFR 1.16(a))</small>	\$ 790
				Total of above Calculations =	
	Reduction by 50% for filing by small entity (Note 37 CFR 1.9, 1.27, 1.28) =				\$
	Assignment Recordation Fee =				\$
	TOTAL =				\$ 1,010.00
<p>1. Small entity status:</p> <p>a. <input type="checkbox"/> A small entity statement is enclosed.</p> <p>b. <input type="checkbox"/> A small entity statement was filed in the prior non-provisional application and such status is still proper and desired.</p> <p>c. <input type="checkbox"/> Is no longer claimed.</p> <p>2. <input checked="" type="checkbox"/> A general authorization is hereby granted to charge deposit account number 08-0380 for any fees required under 37 CFR 1.16 and 1.17 in order to maintain pendency of this application. A copy of this authorization is enclosed for accounting purposes.</p> <p>3. <input checked="" type="checkbox"/> A check is enclosed for \$ <u>1,010.00</u> <span style="margin-left: 100px;"><input type="checkbox"/> Please charge \$ _____ to Deposit Account No. 08-0380.</span></p> <p>4. <input type="checkbox"/> Other: _____</p>					
Submitted by Typed or Printed Name		Anne J. Collins		Reg. Number	40,564
Signature		<i>Anne J. Collins</i>		Date	<i>January 2, 1998</i>

-1-

Date: 1-2-98 Express Mail Label No. EM361199084US

Inventors: Gerd Sutter, Marion Ohlmann and  
Volker Erfle  
Attorney's Docket No.: GSF97-06

## RECOMBINANT MVA VIRUS, AND THE USE THEREOF

### RELATED APPLICATIONS

This is a continuation application of PCT/EP96/02926  
filed July 3, 1996 which claims priority to Danish patent  
5 application DK 0782/95 filed July 4, 1995. The contents of  
PCT/EP96/02926 and DK 0782/95 are incorporated herein by  
reference in their entirety.

### BACKGROUND OF THE INVENTION

Vaccinia virus, a member of the genus Orthopoxvirus in  
10 the family of Poxviridae, was used as live vaccine to  
immunize against the human smallpox disease. Successful  
world-wide vaccination with vaccinia virus culminated in  
the eradication of variola virus, the causative agent of  
the smallpox (The global eradication of smallpox. Final  
15 report of the global commission for the certification of  
smallpox eradication. History of Public Health, No.4,

-2-

Geneva: World Health Organization, 1980). Since that WHO declaration, vaccination has been universally discontinued except for people at high risk of poxvirus infections (e.g. laboratory workers).

5 More recently, vaccinia viruses have also been used to engineer viral vectors for recombinant gene expression and for the potential use as recombinant live vaccines (Mackett, M. et al., *P.N.A.S. USA*, 79:7415-7419 (1982); Smith, G.L et al., *Biotech. and Genetic Engineering Reviews*  
10 2:383-407, (1984)). This entails DNA sequences (genes) which code for foreign antigens being introduced, with the aid of DNA recombination techniques, into the genome of the vaccinia viruses. If the gene is integrated at a site in the viral DNA which is non-essential for the life cycle of  
15 the virus, it is possible for the newly produced recombinant vaccinia virus to be infectious, that is to say able to infect foreign cells and thus to express the integrated DNA sequence (EP Patent Applications No. 83,286 and No. 110,385)). The recombinant vaccinia viruses  
20 prepared in this way can be used, on the one hand, as live vaccines for the prophylaxis of infectious diseases, on the other hand, for the preparation of heterologous proteins in eukaryotic cells.

Recombinant vaccinia virus expressing the  
25 bacteriophage T7 RNA polymerase gene allowed the establishment of widely applicable expression systems for the synthesis of recombinant proteins in mammalian cells

-3-

(Moss, B., et al., *Nature*, 348:91-92 (1990)). In all protocols, recombinant gene expression relies on the synthesis of the T7 RNA polymerase in the cytoplasm of eukaryotic cells. Most popular became a protocol for  
5 transient-expression (Fuerst, T.R., et al., *Proc. Natl. Acad. Sci. USA*, 83:8122-8126 (1986) and US patent application 7,648,971)). First, a foreign gene of interest is inserted into a plasmid under the control of the T7 RNA polymerase promoter. In the following, this plasmid is  
10 introduced into the cytoplasm of cells infected with a recombinant vaccinia virus producing T7 RNA polymerase using standard transfection procedures.

This transfection protocol is simple because no new recombinant viruses need to be made and very efficient with  
15 greater than 80% of the cells expressing the gene of interest (Elroy-Stein, O. and Moss, B., *Proc. Natl. Acad. Sci. USA*, 87:6743-6747 (1990)). The advantage of the vaccinia virus/T7 RNA polymerase hybrid system over other transient expression systems is very likely its  
20 independence on the transport of plasmids to the cellular nucleus. In the past, the system has been extremely useful for analytical purposes in virology and cell biology (Buonocore, L. and Rose, J.K, *Nature*, 345:625-628, (1990); Pattnaik, A.K and Wertz, G.W., *Proc. Natl. Acad. Sci. USA*,  
25 88:1379-1383 (1991); Karschin, A. et al., *FEBS Lett.* 278: 229-233 (1991), Ho, B.Y. et al., *FEBS Lett.*, 301:303-306 (1992); Buchholz, C.J. et al., *Virology*, 204:770-776

-4-

(1994)). However, important future applications of the vaccinia virus/T7 RNA polymerase hybrid system, as e.g. to generate recombinant proteins or recombinant viral particles for novel therapeutic or prophylactic approaches in humans, might be hindered by the productive replication of the recombinant vaccinia vector.

Vaccinia virus is infectious for humans and upon vaccination during the smallpox eradication campaign occasional serious complications were observed. The best overview about the incidence of complications is given by a national survey in the United States monitoring vaccination of about 12 million people with a vaccine based on the New York City Board of Health strain of vaccinia virus (Lane, J. et al. *New Engl. J. Med.*, 281:1201-1208, (1969)). Therefore the most exciting possibility to use vaccinia virus as vector for the development of recombinant live vaccines has been affected by safety concerns and regulations. Furthermore, most of the recombinant vaccinia viruses described in the literature are based on the Western Reserve strain of vaccinia virus. On the other hand, it is known that this strain has a high neurovirulence and is thus poorly suited for use in humans and animals (Morita et al., *Vaccine*, 5:65-70 (1987)).

For vector applications health risks would be lessened by the use of a highly attenuated vaccinia virus strain. Several such strains of vaccinia virus were especially developed to avoid undesired side effects of smallpox

vaccination. Thus, the modified vaccinia virus Ankara (MVA) has been generated by long-term serial passages of the Ankara strain of vaccinia virus (CVA) on chicken embryo fibroblasts (for review see Mayr, A., et al., *Infection*, 5 3:6-14 (1975); Swiss Patent No. 568,392)). The MVA virus was deposited in compliance with the requirements of the Budapest Treaty at CNCM (Institut Pasteur, Collection Nationale de Cultures Microorganisms, 25, rue du Docteur Roux, 75724 Paris Cedex 15) on December 15, 1987 under  
10 Depositary No. I-721. MVA is distinguished by its great attenuation, that is to say by diminished virulence or infectiosity while maintaining good immunogenicity. The MVA virus has been analyzed to determine alterations in the genome relative to the wild CVA strain. Six major  
15 deletions of genomic DNA (deletion I, II, III, IV, V, and VI ) totaling 31,000 base pairs have been identified (Meyer, H., et al., *J. Gen. Virol.* 72:1031-1038 (1991)). The resulting MVA virus became severely host cell restricted to avian cells.

20 Furthermore, MVA is characterized by its extreme attenuation. When tested in a variety of animal models, MVA was proven to be avirulent even in immunosuppressed animals. More importantly, the excellent properties of the MVA strain have been demonstrated in extensive clinical  
25 trials (Mayr et al., *Zbl. Bakt. Hyg. I, Abt. Org. B* 167: 375-390 (1987), Stickl et al., *Dtsch. med. Wschr.* 99: 2386-2392 (1974)). During these studies in over 120,000

-6-

humans, including high risk patients, no side effects were associated with the use of MVA vaccine.

MVA replication in human cells was found to be blocked late in infection preventing the assembly to mature  
5 infectious virions. Nevertheless, MVA was able to express viral and recombinant genes at high levels even in non-permissive cells and was proposed to serve as an efficient and exceptionally safe gene expression vector (Sutter, G. and Moss, B., *Proc. Natl. Acad. Sci. USA*  
10 89:10847-10851 (1992)). Recently, novel vaccinia vector systems were established on the basis of MVA, having foreign DNA sequences inserted at the site of deletion III within the MVA genome or within the TK gene (Sutter, G. and Moss, B. *Dev. Biol. Stand. Basel*, Karger 84:195-200  
15 (1995) and US patent 5,185,146)).

To further exploit the use of MVA a novel possible way to introduce foreign genes by DNA recombination into the MVA strain of vaccinia virus has been sought. Since the intention was not to alter the genome of the MVA virus, it  
20 was necessary to use a method which complied with this requirement. According to the present invention a foreign DNA sequence was recombined into the viral DNA precisely at the site of a naturally occurring deletion in the MVA genome.



-7-

## SUMMARY OF THE INVENTION

The present invention thus, inter alia, comprises the following, alone or in combination:

- A recombinant MVA virus containing and capable of
- 5 expressing at least one foreign gene inserted at the site of a naturally occurring deletion within the MVA genome;
- a recombinant MVA virus as above containing and capable of expressing at least one foreign gene inserted at the site of deletion II within the MVA genome;
- 10 a recombinant MVA virus as above wherein the foreign gene codes for a marker, a therapeutic gene or an antigenic determinant;
- a recombinant MVA virus as above wherein the foreign gene codes for an antigenic determinant from a pathogenic
- 15 virus, a bacteria, or other microorganism, or from a parasite, or a tumor cell;
- a recombinant MVA virus as above wherein the foreign gene codes for an antigenic determinant from Plasmodium Falciparum, Mycobacteria, Herpes virus, influenza virus,
- 20 hepatitis, or human immunodeficiency viruses.
- a recombinant MVA virus as above wherein the antigenic determinant is HIV nef or human tyrosinase;
- a recombinant MVA virus as above which is MVA-LAI<sub>nef</sub> or MVA-hTYR;
- 25 a recombinant MVA virus as above wherein the foreign gene codes for T7 RNA polymerase;
- a recombinant MVA virus as above which is MVA-T7 pol;

-8-

a recombinant MVA virus as above wherein the foreign gene is under transcriptional control of the vaccinia virus early/late promoter P7.5;

recombinant MVA viruses as above essentially free from  
5 viruses being able to replicate in human cells;

the use of a recombinant MVA virus as above for the transcription of DNA sequences under transcriptional control of a T7 RNA polymerase promoter;

a eukaryotic cell infected by a recombinant MVA virus  
10 as any above;

a cell infected by a recombinant MVA virus as above wherein the foreign gene code for T7 RNA polymerase;

a cell infected by a recombinant MVA virus as above wherein the foreign gene code for T7 RNA polymerase,  
15 additionally containing one or more expression vectors carrying one or more foreign genes under transcriptional control of a T7 RNA polymerase promoter;

the use of cells as above for the production of the polypeptides encoded by said foreign genes comprising:

- 20 a) culturing said cells under suitable conditions,  
and  
b) isolating the polypeptides encoded by said foreign genes.

a cell infected by a recombinant MVA virus as above  
25 wherein the foreign gene code for T7 RNA polymerase, additionally containing expression vectors carrying viral genes, and/or a viral vector construct encoding the genome

of a viral vector under transcriptional control of a T7 RNA polymerase promoter;

the use of a cells as above for the production viral particles comprising:

5           a)     culturing said cells under suitable conditions,  
                  and

          b)     isolating the viral particles;

          a cell infected by a recombinant MVA virus as above wherein the foreign gene code for T7 RNA polymerase,

10          additionally containing

          a)     an expression vector carrying a retroviral vector construct capable of infecting and directing the expression in target cells of one or more foreign genes carried by said retroviral vector  
15                   construct, and

          b)     one or more expression vectors carrying the genes encoding the polypeptides required for the genome of said retroviral vector construct to be  
20                   packaged under transcriptional control of a T7

          RNA polymerase promoter;

the use of cells as above for the production of retroviral particles comprising

          a)     culturing said cells under suitable conditions,  
                  and

25          b)     isolating the retroviral particles;

-10-

a vaccine containing a recombinant MVA virus as above wherein the foreign gene code for an antigenic determinant in a physiologically acceptable carrier;

the use of a recombinant MVA virus as above wherein  
5 the foreign gene code for an antigenic determinant preparation of a vaccine;

the use of a vaccine as above for the immunization of a living animal body, including a human;

the use of a vaccine as above containing MVA-LAInef  
10 for the prevention or treatment of HIV infection or AIDS;

the use of a vaccine as above containing MVA-hTYR for the prevention or treatment of melanomas;

a vaccine comprising as a first component, a recombinant MVA virus as above wherein the foreign gene  
15 code for T7 RNA polymerase in a physiologically acceptable carrier, and as a second component a DNA sequence carrying an antigenic determinant under transcriptional control of a T7 RNA polymerase promoter in a physiologically acceptable carrier, the two components being contained together or  
20 separate;

the use of a vaccine as above for the immunization of a living animal body, including a human, comprising inoculation of said living animal body, including a human, with the first and second component of the vaccine either  
25 simultaneously or with a timelag using the same inoculation site; and

-11-

The term "gene" means any DNA sequence which codes for a protein or peptide.

The term "foreign gene" means a gene inserted in a DNA sequence in which it is not normally found.

5       The foreign gene can be a marker gene, a therapeutic gene, a gene encoding an antigenic determinant, or a viral gene, for example. Such genes are well known in the art.

#### BRIEF DESCRIPTION OF THE FIGURES

10       Figure 1 is a schematic map of the genome of MVA and plasmid for insertion of foreign DNA by homologous recombination: HindIII restriction sites within the genome of MVA are indicated at the top; the 900-bp HindIII-HindIII N fragment that overlaps the junction of deletion II within the MVA genome is shown; MVA DNA sequences adjacent to  
15       deletion II (flank 1 and flank 2) were amplified by PCR and used for the construction of insertion plasmid pUC II LZ.

      Figure 2 is a schematic map of pUC II LZ P7.5: MVA vector plasmid for insertion into deletion II containing P11-LacZ expression cassette and the vaccinia virus  
20       early/late promoter P7.5 to express genes of interest that can be cloned into the *Sma*I site of the plasmid.

      Figure 3 is a schematic map of pUCII LZdel P7.5: MVA vector plasmid for insertion of foreign genes at the site of deletion II in the MVA genome, containing a  
25       self-deleting P11-LacZ expression cassette and the vaccinia virus early/late promoter P7.5 to express genes of interest

-12-

that can be cloned into the *Sma*I/*Not*I cloning site of the plasmid.

Figure 4 is a schematic map of the construction of recombinant virus MVA-T7pol: schematic maps of the MVA genome (*Hind*III restriction endonuclease sites) and the vector plasmid pUC II LZ T7pol that allows insertion of the T7 RNA polymerase gene at the site of deletion II within the *Hind*III N fragment of the MVA genome.

Figure 5 is a schematic map of the construction of MVA-LAInef: schematic maps of the MVA genome (*Hind*III restriction endonuclease sites) and the vector plasmid pUC II LZdel P7.5-LAInef that allows insertion of the nef gene of HIV-1 LAI at the site of deletion II within the *Hind*III N fragment of the MVA genome.

Figure 6 is a schematic map of the construction of MVA-hTYR: schematic maps of the MVA genome (*Hind*III restriction endonuclease sites) and the vector plasmid pUC II LZdel P7.5-TYR that allows insertion of the human tyrosinase gene at the site of deletion II within the *Hind*III N fragment of the MVA genome.

#### DETAILED DESCRIPTION OF THE INVENTION

It is an object of the present invention to provide a recombinant MVA virus which can serve as an efficient and exceptionally safe expression vector.

Another object of the present invention is to provide a simple, efficient and safe method for the production of

-13-

polypeptides, e.g. antigens or therapeutic agents, recombinant viruses for vaccines and viral vectors for gene therapy.

Still another object of the present invention is to  
5 provide an expression system based on a recombinant MVA virus expressing T7 RNA polymerase, and methods for the production of polypeptides, e.g. antigens or therapeutic agents, or for generating viral vectors for gene therapy or vaccines, based on this expression system.

#### 10 The present Invention

Modified vaccinia virus Ankara (MVA), a host range restricted and highly attenuated vaccinia virus strain, is unable to multiply in human and most other mammalian cell lines tested. But since viral gene expression is  
15 unimpaired in non-permissive cells the recombinant MVA viruses according to the invention may be used as exceptionally safe and efficient expression vectors.

#### RECOMBINANT MVA VIRUSES ENCODING AN ANTIGENIC DETERMINANT

In one embodiment, the present invention relates to  
20 recombinant MVA vaccinia viruses which contain a gene which codes for a foreign antigen, preferably of a pathogenic agent, and vaccines containing such a virus in a physiologically acceptable form. The invention also relates to methods for the preparation of such recombinant  
25 MVA vaccinia viruses or vaccines, and to the use of these

vaccines for the prophylaxis of infections caused by such pathogenic agents.

In a preferred embodiment of the invention, the foreign gene inserted in the MVA virus is a gene encoding  
5 HIV nef.

We have constructed recombinant MVA viruses that allow expression of the HIV-1 nef gene under the control of the vaccinia virus early/late promoter P7.5. The regulatory Nef protein of primate lentiviruses is synthesized early in  
10 the viral replication cycle and has been shown to be essential for high titer virus replication and disease induction *in vivo*. This suggests that HIV Nef might play a crucial role in AIDS pathogenesis. The molecular mechanism(s) by which Nef contributes to increased viral  
15 infectivity and to HIV pathogenicity need to be further elucidated. However, Nef is immunogenic and Nef-specific antigen can be used as a vaccine against HIV infection and AIDS.

In this context, the recombinant MVA virus expressing  
20 the HIV nef gene can be used for immunization of human beings, on one hand, as a prophylactic vaccine against human HIV, and on the other hand, for immunotherapy of HIV infected or AIDS patients. Furthermore, the recombinant MVA virus expressing the HIV nef gene can be used for the  
25 production of recombinant HIV Nef protein.



In another preferred embodiment of the invention the foreign gene inserted in the MVA virus is a gene encoding human tyrosinase.

We have constructed recombinant MVA viruses that allow  
5 expression of the human tyrosinase gene under the control  
of the vaccinia virus early/late promoter P7.5. Recently,  
human tyrosinase was identified as a melanoma-specific  
tumor antigen that allows generation of anti-tumor  
cytolytic T-lymphocytes (Beichard, V., et al., *J. Exp.*  
10 *Med.*, 178:489-495 (1993)). Since among normal cells, only  
melanocytes appear to express the tyrosinase gene,  
tyrosinase is a useful target antigen for immunotherapy of  
melanomas. Therefore, the recombinant MVA virus expressing  
the human tyrosinase gene can be used in melanoma patients  
15 to induce immune responses that provoke tumor rejection or  
prevent metastasis. Recombinant MVA virus expressing the  
human tyrosinase gene can be used directly as an  
anti-melanoma vaccine, or the virus can be used to prepare  
anti-melanoma vaccines. In one example, the recombinant  
20 MVA virus expressing the human tyrosinase gene can be used  
for the production of recombinant tyrosinase protein which  
is used as antigen in vaccine preparations. In another  
example, using the recombinant MVA virus expressing the  
human tyrosinase gene as expression vector, cells derived  
25 from a tumor patient can be modified *in vitro* to express  
tyrosinase and then transferred back to the patient to  
induce anti-tumor immune responses. A vaccine prepared on

-16-

the basis of recombinant MVA expressing the human tyrosinase gene can be used either parenterally or locally at the site of the tumor. To prevent tumor metastasis or to phenotypically change the tumor e.g. in size, shape, consistency, vascularization or other features. A vaccine prepared on the basis of recombinant MVA expressing the human tyrosinase gene can be used before, during, or after surgical extirpation of the tumor.

For the preparation of vaccines, the MVA vaccinia viruses according to the invention are converted into a physiologically acceptable form. This can be done based on the experience in the preparation of MVA vaccines used for vaccination against smallpox (as described by Stickl, H. et al., *Dtsch. med. Wschr.* 99:2386-2392 (1974)). Typically, about  $10^6$ - $10^8$  particles of the recombinant MVA are freeze-dried in 100 ml of phosphate-buffered saline (PBS) in the presence of 2% peptone and 1% human albumin in an ampoule, preferably a glass ampoule. The lyophilisate can contain extenders (such as mannitol, dextran, sugar, glycine, lactose or polyvinylpyrrolidone) or other aids (such as antioxidants, stabilizers, etc.) suitable for parenteral administration. The glass ampoule is then sealed and can be stored, preferably at temperatures below  $-20^{\circ}\text{C}$ ., for several months.

For vaccination or therapy the lyophilisate can be dissolved in 0.1 to 0.5 ml of an aqueous solution, preferably physiological saline, and administered either

-17-

parenterally, for example by intramuscular inoculation or locally, for example by inoculation into a tumor or at the site of a tumor. Vaccines or therapeutics according to the invention are preferably injected intramuscularly (Mayr, A. et al., *Zbl. Bakt. Hyg., I. Abt. Orig. B* 167:375-390 (1978)). The mode of administration, the dose and the number of administrations can be optimized by those skilled in the art in a known manner. It is expedient where appropriate to administer the vaccine several times over a lengthy period in order to obtain appropriate immune responses against the foreign antigen.

#### THE USE OF RECOMBINANT MVA VIRUSES FOR THE PRODUCTION OF HETEROLOGOUS POLYPEPTIDES

The recombinant MVA vaccinia viruses according to the invention can also be used to prepare heterologous polypeptides in eukaryotic cells. This entails cells being infected with the recombinant vaccinia viruses. The gene which codes for the foreign polypeptide is expressed in the cells, and the expressed heterologous polypeptide is isolated. The methods to be used for the production of such heterologous polypeptides are generally known to those skilled in the art (EP-A-206,920 and EP-A-205,939). The polypeptides produced with the aid of the recombinant MVA viruses are, by reason of the special properties of the MVA viruses, more suitable for use as medicaments in humans and animals.

-18-

RECOMBINANT MVA VIRUSES ENCODING T7 RNA POLYMERASE AND THE  
USE THEREOF FOR THE EXPRESSION OF DNA SEQUENCES UNDER  
TRANSCRIPTIONAL CONTROL OF A T7 RNA POLYMERASE PROMOTER

In a further embodiment of the present invention we  
5 have constructed recombinant MVA viruses that allow  
expression of the bacteriophage T7 RNA polymerase gene  
under the control of the vaccinia virus early/late promoter  
P7.5. The usefulness of MVA-T7pol recombinant viruses as  
expression system has been tested in transient transfection  
10 assays to induce expression of recombinant genes under the  
control of a T7 RNA polymerase promoter. Using the *E. coli*  
chloramphenicol acetyltransferase (CAT) gene as a reporter  
gene we found that MVA-T7pol induced CAT gene expression as  
effectively as a vaccinia/ T7pol recombinant virus derived  
15 from the replication-competent WR strain of vaccinia virus.

The MVA/T7 polymerase hybrid system according to the  
invention can thus be used as a simple, efficient and safe  
mammalian expression system for production of polypeptides  
in the absence of productive vaccinia virus replication.

20 This expression system can also be used for generating  
recombinant viral particles for vaccination or gene therapy  
by transformation of cell lines infected with recombinant  
MVA expressing T7 RNA polymerase, with DNA-constructs  
containing all or some of the genes, and the genome or  
25 recombinant genome necessary for generating viral  
particles, e.g MVA particles or retroviral particles, under  
transcriptional control of a T7 RNA polymerase promoter.

-19-

Retroviral vector systems consist of two components:

- 1) the retroviral vector itself is a modified retrovirus (vector plasmid) in which the genes encoding for the viral proteins have been replaced by therapeutic genes and marker genes to be transferred to the target cell. Since the replacement of the genes encoding for the viral proteins effectively cripples the virus it must be rescued by the second component in the system which provides the missing viral proteins to the modified retrovirus.

The second component is:

- 2) a cell line that produces large quantities of the viral proteins, however lacks the ability to produce replication competent virus. This cell line is known as the packaging cell line and consists of a cell line transfected with one or more plasmids carrying the genes (genes encoding the gag, pol and env polypeptides) enabling the modified retroviral vector to be packaged.

To generate the packaged vector, the vector plasmid is transfected into the packaging cell line. Under these conditions the modified retroviral genome including the inserted therapeutic and marker genes is transcribed from

-20-

the vector plasmid and packaged into the modified retroviral particles (recombinant viral particles). This recombinant virus is then used to infect target cells in which the vector genome and any carried marker or  
5 therapeutic genes becomes integrated into the target cell's DNA. A cell infected with such a recombinant viral particle cannot produce new vector virus since no viral proteins are present in these cells. However, the DNA of the vector carrying the therapeutic and marker genes is  
10 integrated in the cell's DNA and can now be expressed in the infected cell.

The recombinant MVA virus according to the invention expressing T7 RNA polymerase can be used to produce the proteins required for packaging retroviral vectors. To do  
15 this the gag, pol and env genes of a retrovirus (e.g. the Murine Leukemia Virus (MLV)) are placed under transcriptional control of a T7 RNA polymerase promoter in one or more expression vectors (e.g. plasmids). The expression vectors are then introduced into cells infected  
20 with the recombinant MVA virus expressing T7 RNA polymerase, together with an expression vector carrying a retroviral vector construct, possibly under transcriptional control of a T7 RNA polymerase promoter.

WO 94/29437, WO 89/11539 and WO 96/7748 describes  
25 different types of retroviral vector which can be packaged using the packaging system described above.

A further use of the recombinant MVA virus expressing T7 RNA polymerase is to generate recombinant proteins, noninfectious virus particles, or infectious mutant virus particles for the production of vaccines or therapeutics (Buchholz et al., *Virology*, 204:770-776 (1994) and EP-B1-1356695)). To do this viral genes (e.g. the gag-pol and env genes of HIV-1) are placed under transcriptional control of the T7 promoter in an expression vector (e.g. plasmid or another recombinant MVA virus). This construct is then introduced into cells infected with the recombinant MVA virus expressing T7 RNA polymerase. The recombinant viral genes are transcribed with high efficiency, recombinant proteins are made in high amounts and can be purified. Additionally, expressed recombinant viral proteins (e.g., HIV-1 env, gag) may assemble to viral pseudo-particles that budd from the cells and can be isolated from the tissue culture medium. In another embodiment, viral proteins (from e.g. HIV, SIV, Measles virus) expressed by the MVA-T7 pol system may rescue an additionally introduced mutant virus (derived from e.g. HIV, SIV, Measles virus) by overcoming a defect in attachment and infection, uncoating, nucleic acid replication, viral gene expression, assembly, budding or another step in viral multiplication to allow production and purification of the mentioned mutant virus.

MVA-T7pol can also be used together with DNA sequences carrying the gene of an antigen of interest (e.g. the gene

-22-

of HIV, nef, tat, gag, pol , or env or others) for immunization. First, a coding sequence of a given antigen (e.g HIV, HCV, HPV, HSV, measles virus, influenza virus or other) are cloned under control of a T7 RNA polymerase promoter preferably in a plasmid vector and the resulting DNA construct is amplified and purified using standard laboratory procedures. Secondly, the vector DNA is inoculated simultaneously or with appropriate limelags together with MVA-T7pol. At the site of inoculation the recombinant gene of interest is expressed transiently in cells containing both the vector DNA and MVA-T7 pol and the corresponding antigen is presented to the host immune system stimulating an antigen-specific immune response. This protocol using the non-replication vaccinia with MVA -T7 pol represents a promising novel approach to nucleic acid vaccination allowing efficient transient expression of a given antigen, but avoiding the potential risk of constitutive gene expression.

THE RECOMBINANT MVA VACCINIA VIRUSES CAN BE PREPARED AS SET  
OUT HEREINAFTER

A DNA-construct which contains a DNA-sequence which codes for a foreign polypeptide flanked by MVA DNA sequences adjacent to a naturally occurring deletion, e.g. deletion II, within the MVA genome, is introduced into cells infected with MVA, to allow homologous recombination.



Once the DNA-construct has been introduced into the eukaryotic cell and the foreign DNA has recombined with the viral DNA, it is possible to isolate the desired recombinant vaccinia virus in a manner known per se,

5 preferably with the aid of a marker (compare Nakano *et al.*, *Proc. Natl. Acad. Sci. USA*, 79:1593-1596 (1982); Franke *et al.*, *Mol. Cell. Biol.*, 1918-1924 (1985); Chakrabarfi *et al.*, *Mol. Cell. Biol.*, 3403-3409 (1985); Fathi *et al.*, *Virology* 97-105 (1986)).

10 The DNA-construct to be inserted can be linear or circular. A circular DNA is preferred, especially a plasmid. The DNA-construct contains sequences flanking the left and the right side of a naturally occurring deletion, e.g. deletion II, within the MVA genome (Altenburger, W.,  
15 Suter, C.P. and Altenburger J., *Arch. Virol.*, 105:15-27 (1989)). The foreign DNA sequence is inserted between the sequences flanking the naturally occurring deletion. The foreign DNA sequence can be a gene coding for a therapeutic polypeptide, e.g. t-PA or interferon, or an antigenic  
20 determinant from a pathogenic agent. Pathogenic agents can be viruses, bacteria and parasites which may cause a disease, as well as tumor cells which multiply unrestrictedly in an organism and may thus lead to pathological growths. Examples of such pathogenic agents  
25 are described in Davis, B.D. *et al.*, (Microbiology, 3rd ed., Harper international Edition). Preferred antigens of pathogenic agents are those of human immunodeficiency

viruses (e.g. HIV-1 and HIV-2), of mycobacteria causing tuberculosis, of the parasite *Plasmodium Falciparum*, and of melanoma cells.

For the expression of a DNA sequence or gene, it is  
5 necessary for regulatory sequences, which are required for  
the transcription of the gene, to be present on the DNA.  
Such regulatory sequences (called promoters) are known to  
those skilled in the art, and includes for example those of  
the vaccinia 11 kDa gene as are described in EP-A-198,328,  
10 and those of the 7.5 kDa gene (EP-A-110,385).

The DNA-construct can be introduced into the MVA  
infected cells by transfection, for example by means of  
calcium phosphate precipitation (Graham et al., *Virology*,  
52:456-467 (1973); Wigler et al., *Cell* 777-785 (1979)) by  
15 means of electroporation (Neumann et al., *EMBO J.*, 1:841-  
845 (1982)), by microinjection (Graessmann et al., *Meth.*  
*Enzymol.* 101:482-492 (1983)), by means of liposomes  
(Straubinger et al., *Methods in Enzymology*, 101:512-527  
(1983)), by means of spheroplasts (Schaffner, *Proc. Natl.*  
20 *Acad. Sci. USA*, 77:2163- 2167 (1980)) or by other methods  
known to those skilled in the art. Transfection by means of  
calcium phosphate precipitation is preferred.

The detailed examples which follow are intended to  
contribute to a better understanding of the present  
25 invention. However, it is not intended to give the  
impression that the invention is confined to the  
subject-matter of the examples.

-25-

## EXAMPLES

## 1. Growing and purification of the viruses

## 1.1 Growing of the MVA virus

The MVA virus is a highly attenuated vaccinia virus derived from the vaccinia virus strain Ankara (CVA) by long-term serial passages on primary chicken embryo fibroblast (CEF) cultures. For a general review of the history of the production, the properties and the use of MVA strain, reference may be made to the summary published by Mayr et al., in *Infection*, 3:6-14 (1975). Due to the attenuation in CEF, the MVA virus replicates to high titers in this avian host cell. In mammalian cells, however, MVA is severely growth restricted, and typical plaque formation by the virus is not detectable. Therefore, MVA virus was grown on CEF cells. To prepare CEF cells, 11 - day old embryos were isolated from incubated chicken eggs, the extremities are removed, and the embryos are minced and dissociated in a solution composed of 0.25% trypsin at 37°C for 20 minutes. The resulting cell suspension was filtered and cells were sedimented by centrifugation at 2000 rpm in a Sorvall RC-3B centrifuge at room temperature for 5 minutes, resuspended in 10 volumes of medium A (MEM Eagle, for example obtainable from Life Technologies GmbH, Eggenstein, Germany), and sedimented again by centrifugation at 2000 rpm in a Sorvall RC-3B centrifuge at room temperature for 5 minutes. The cell pellet was

-26-

reconstituted in medium A containing 10% fetal calf serum (FCS), penicillin (100 units/ml), streptomycin (100 mg/ml) and 2 mM glutamine to obtain a cell suspension containing 500,000 cells/ml. CEF cells obtained in this way were

5 spread on cell culture dishes. They were left to grow in medium A in a CO<sub>2</sub> incubator at 37°C for 1-2 days, depending on the desired cell density, and were used for infection either directly or after one further cell passage. A detailed description of the preparation of primary cultures

10 can be found in the book by R.I. Freshney, "Culture of animal cell, Alan R. Liss Verlag, New York (1983) Chapter 11, page 99 et seq.

MVA viruses were used for infection as follows. CEF cells were cultured in 175 cm<sup>2</sup> cell culture bottles. At

15 90-100% confluence, the medium was removed and the cells were incubated for one hour with an MVA virus suspension (0.01 infectious units (IU) per cell, 0.02 ml/cm<sup>2</sup>) in medium A. Then more medium A was added (0.2 ml/cm<sup>2</sup>) and the bottles were incubated at 37°C for 2-3 days (until about

20 90% of the cells show cytopathogenic effect). Crude virus stocks were prepared by scraping cell monolayers into the medium and pelleting the cell material by centrifugation at 3000 rpm in a Sorvall RC-3B centrifuge at 4°C for 5 minutes. The crude virus preparation was stored at -20°C

25 before processing (e.g., virus purification).

-27-

## 1.2 Purification of the viruses

The purification steps undertaken to obtain a virus preparation which was as pure as possible and free from components specific to the host cell were similar to those described by Joklik, *Virology*, 18:9-18 (1962)). Crude virus stocks which had been stored at  $-20^{\circ}\text{C}$  were thawed and suspended once in PBS (10-20 times the volume of the sediment), and the suspension was centrifuged as above. The new sediment was suspended in 10 times the volume of Tris buffer 1 (10 mM Tris-HCl pH 9.0,), and the suspension was briefly treated with ultrasound (Labsonic L, B.Braun Biotech International, Melsungen Germany; 2x10 seconds at 60 watts and room temperature) in order to further disintegrate cell debris and to liberate the virus particles from the cellular material. The cell nuclei and the larger cell debris were removed in the subsequent brief centrifugation of the suspension (Sorvall GSA rotor obtainable from DuPont Co., D-6353 Bad Nauheim, FRG; 3 minutes at 3000 rpm and  $10^{\circ}\text{C}$ .). The sediment was once again suspended in Tris buffer 1, treated with ultrasound and centrifuged, as described above. The collected supernatants containing the free virus particles were combined and layered over a cushion of 10 ml of 36% sucrose in 10 mM Tris-HCl, pH 9.0, and centrifuged in a Beckman SW 27/SW 28 rotor for 80 minutes with 13,500 rpm at  $40^{\circ}\text{C}$ . The supernatant was discarded, and the sediment containing the virus particles was taken up in 10 ml of 1 mM Tris-HCl, pH

-28-

9.0, homogenized by brief treatment with ultrasound (2x10 seconds at room temperature, apparatus as described above), and applied to a 20-40% sucrose gradient (sucrose in 1 mM Tris-HCl, pH 9.0) for further purification. The gradient  
5 was centrifuged in Beckmann SW41 rotor at 13,000 rpm for 50 minutes at 4°C. After centrifugation, discrete bands containing virus particles were harvested by pipetting after aspirating volume above band. The obtained sucrose solution was diluted with three volumes PBS and the virus  
10 particles were sedimented again by centrifugation (Beckmann SW 27/28, 60 minutes at 13,500 rpm, 4°C.). The pellet, which now consisted mostly of pure virus particles, was resuspended in PBS and equilibrated to virus concentrations corresponding on average to  $1-5 \times 10^9$  IU/ml. The purified  
15 virus stock solution was stored at -80°C and used either directly or diluted with PBS for subsequent experiments.

### 1.3 Cloning of MVA virus

To generate homogeneous stock virus preparations MVA virus obtained from Prof. Anton Mayr was cloned by limiting  
20 dilution during three consecutive passages in CEF cultured on 96-well tissue culture plates. The MVA clone F6 was selected and amplified in CEF to obtain working stocks of virus that served as starting material for the generation of recombinant MVA viruses described in this patent  
25 application as well as for the generation of recombinant MVA viruses described previously (Sutter, G. and Moss, B.,

*Proc. Natl. Acad. Sci. USA* , 89:10847-10851 (1992); Sutter, G. et al., *Vaccine*, 12:1032-1040 (1994); Hirsch, V. et al., *J. Virol.*, 70:3741-3752 (1996)).

## 2. Construction and characterization of recombinant MVA 5 viruses

### 2.1. Construction of vector plasmids

To allow the generation of recombinant MVA viruses novel vector plasmids were constructed. Insertion of foreign genes into the MVA genome was targeted precisely to  
10 the site of the naturally occurring deletion II in the MVA genome. Sequences of MVA DNA flanking the site of a 2500-bp deletion in the *HindIII* N fragment of the MVA genome (Altenburger, W. et al., *J. Arch. Virol.*, 105:15-27 (1989)) were amplified by PCR and cloned into the multiple  
15 cloning site of plasmid pUC18. The primers for the left 600-bp DNA flank were 5'-CAG CAG GGT ACC CTC ATC GTA CAG GAC GTT CTC-3' (SEQ ID NO: 1) and 5'-CAG CAG CCC GGG- TAT TCG ATG ATT ATT TTT AAC AAA ATA ACA-3' (SEQ ID NO: 2) (sites for restriction enzymes *KpnI* and *SmaI* are  
20 underlined).

The primers for the right 550-bp DNA flank were 5'-CAG CAG CTG CAG GAA TCA TCC ATT CCA CTG AAT AGC-3' (SEQ ID NO: 3); and 5'-CAG CAG GCA TGC CGA CGA ACA AGG AAC TGT AGC AGA-3' (SEQ ID NO: 4) (sites for restriction enzymes *PstI*  
25 and *SphI* are underlined). Between these flanks of MVA DNA inserted in pUC18, the *Escherichia coli LacZ* gene under

control of the vaccinia virus late promoter P11 (prepared by restriction digest from pIII LZ, Sutter, G. and Moss, B., *PNAS USA* 89:10847-10851 (1992)) was inserted, using the *Bam*HI site, to generate the MVA insertion vector pUCII LZ

5 (Figure 1). In the following, a 289 bp fragment containing the vaccinia virus early-late promoter P7.5 together with a *Sma*I site for cloning (prepared by restriction digest with *Eco*RI and *Xba*I from the plasmid vector pSC11 (Chakrabarti *et al.*, *Mole. Cell. Biol.*, 5:3403-3409 (1985)) was inserted

10 into the *Sma*I site of pUCII LZ to give the MVA vector pUC II LZ P7.5 [Figure 2]. To construct a vector plasmid that allows isolation of recombinant MVA viruses via transient synthesis of the reporter enzyme  $\beta$ -galactosidase a 330 bp DNA fragment from the 3'-end of the *E. coli LacZ* open

15 reading frame was amplified by PCR (primers were 5'-CAG CAG GTC GAC CCC GAC CGC CTT ACT GCC GCC-3' (SEQ ID NO: 5) and 5'-GGG GGG CTG CAG ATG GTA GCG ACC GGC GCT CAG-3' (SEQ ID NO: 6)) and cloned into the *Sal*I and *Pst*I sites of pUC II LZ P7.5 to obtain the MVA vector pUC II LZdel P7.5 (Figure

20 3). Using the *Sma*I site, this vector plasmid can be used to insert DNA sequences encoding a foreign gene under transcriptional control of the vaccinia virus promoter P7.5 into the MVA genome. After the desired recombinant virus has been isolated by screening for expression of

25  $\beta$ -galactosidase activity further propagation of the recombinant virus leads to the self-deletion of the



-31-

reengineered P11-LacZ expression cassette by homologous recombination.

## 2.2. CONSTRUCTION AND CHARACTERIZATION OF RECOMBINANT VIRUS MVA T7pol

5        A 3.1 kbp DNA fragment containing the entire gene of bacteriophage T7 RNA polymerase under control of the vaccinia virus early/late promoter P7.5 was excised with *EcoR*I from plasmid pTF7-3 (Fuerst, T.R. et al., *P.N.A.S. USA*, 83:8122-8126 (1986), modified by incubation with  
10    Klenow DNA polymerase to generate blunt ends, and cloned into a unique *Sma*I restriction site of pUCII LZ to make the plasmid transfer vector pUCII LZ T7pol (Figure 4). As transcriptional regulator for the expression of the T7 RNA polymerase gene the vaccinia virus early/late promoter P7.5  
15    was chosen. Contrary to stronger vaccinia virus late promoters (e.g. P11) this promoter system allows expression of recombinant genes immediately after the infection of target cells. The plasmid pUCII LZ T7pol that directs the insertion of the foreign-genes into the site of deletion II  
20    of the MVA genome was used to generate the recombinant virus MVA T7pol.

      CEF cells infected with MVA at a multiplicity of 0.05 TCID<sub>50</sub> per cell were transfected with DNA of plasmid pUCII LZ T7pol as described previously (Sutter, G, et al.,  
25    *Vaccine*, 12:1032-1040 (1994)). Recombinant MVA virus expressing the T7 RNA polymerase and co-expressing

-32-

$\beta$ -D-galactosidase (MVA P7.5-T7pol) was selected by five consecutive rounds of plaque purification in CEF cells stained with 5-bromo-4-chloro-3-indolyl  $\beta$ -D-galactoside (300  $\mu$ g/ml). Subsequently, recombinant viruses were

5 amplified by infection of CEF monolayers, and the DNA was analyzed by PCR to confirm genetic homogeneity of the virus stock. Southern blot analysis of MVA-T7pol viral DNA demonstrated stable integration of the recombinant genes at the site of deletion II within the MVA genome.

10 To monitor expression of T7 RNA polymerase by recombinant MVA T7pol [ $^{35}$ S]methionine -labeled polypeptides from virus infected tissue culture were analyzed. Monolayers of the monkey kidney cell line CV-1 grown in 12-well plates were infected with virus at a multiplicity

15 of 20 TCID<sub>50</sub> per cell. At 3 to 5 hours after infection, the medium was removed, and the cultures were washed once with 1 ml of methionine free medium. To each well, 0.2 ml of methionine-free medium supplemented with 50  $\mu$ Ci of [ $^{35}$ S]methionine was added and incubated for 30 minutes at

20 37°C. Cytoplasmic extracts of infected cells were prepared by incubating each well in 0.2 ml of 0.5% Nonidet P-40 lysis buffer for 10 min at 37°C and samples were analyzed by SDS-PAGE. The metabolic labeling of the CV-1 cells with MVA T7pol revealed the synthesis of two additional

25 polypeptides (i) a protein of about 116,000 Da representing the *E. coli*  $\beta$ -galactosidase co-expressed to allow the screening for recombinant virus and (ii) a 98,000 Da

protein with the expected size of the bacteriophage T7 RNA polymerase. The large amount of  $\beta$ -galactocidase made by MVA T7pol is remarkable. The results from the *in vivo* labeling experiments demonstrate a very strong expression  
5 of the P11-LacZ gene construct when inserted into the MVA genome at the site of deletion II indicating that recombinant genes in MVA vector viruses might be expressed more efficiently when inserted into this locus of the MVA genome.

10 The usefulness of MVA-T7pol recombinant viruses as expression system in comparison to the WR-T7pol recombinant virus vTF7-3 (Fuerst et al. 1986) was tested by the co-transfection of DNA of a plasmid vector that is derived from pTM1 (Moss, B., et al., *Nature*, 348:91-92 (1990)) and  
15 contains (cloned into the *Nco*I and *Bam*HI sites of the pTM1 multiple cloning site) the *E. coli* chloramphenicol acetyltransferase (CAT) gene under the control of a T7 RNA polymerase promoter (PT<sub>7</sub>). Transfected and infected CV-1 cells were suspended in 0.2 ml of 0.25 M Tris-HCl (pH 7.5).  
20 After three freeze-thaw cycles, the lysates were cleared by centrifugation, the protein content of the supernatants was determined, and samples containing 0.5, 0.25, 0.1  $\mu$ g total protein were assayed for enzyme activity as described by Mackett, M., et al., *J. Virol.*, 49:857-864 (1984). After  
25 autoradiography, labeled spots were quantitated using the Fuji imaging analysis system.

The results demonstrate that by using the highly attenuated vaccinia vector MVA it is possible to exploit the vaccinia virus-T7 RNA polymerase system as efficiently as by using a fully replication-competent vaccinia virus recombinant.

### 2.3. CONSTRUCTION AND CHARACTERIZATION OF RECOMBINANT VIRUS MVA-LAlnef

A 648 bp DNA fragment containing the entire nef gene of HIV-1 LAI was prepared by PCR from plasmid DNA (pTG1166 kindly provided by M.-P. Kieny, Transgene S.A., Strasbourg; PCR primers were 5'-CAG CAG GGA TCC ATG GGT GGC AAG TGG TCA AAA AGT AGT-3' (SEQ ID NO: 7) and 5'-CAG CAG GGA TCC ATG TCA GCA GTT CTT GAA GTA CTC CGG-3' (SEQ ID NO: 5)), digested with restriction endonuclease *Bam*HI, modified by incubation with Klenow DNA polymerase to generate blunt ends, and cloned into the *Sma*I site of pUC II LZdel P7.5 to make the vector pUC II LZdel P7.5-LAlnef (Figure 5). This plasmid could be used to engineer MVA recombinant virus that expresses the nef gene of HIV-1 LAI under control of the vaccinia virus early/late promoter P7.5.

CEF cells infected with MVA at a multiplicity of 0.05 TCID<sub>50</sub> per cell were transfected with DNA of plasmid pUC II LZdel P7.5-LAlnef as described previously (Sutter, G. et al., *Vaccine*, 12:1032-1040 (1994)). Recombinant MVA viruses containing the nef gene and transiently co-expressing the *E. coli* LacZ marker gene were selected by

-35-

consecutive rounds of plaque purification in CEF cells stained with 5-bromo-4-chloro-3-indolyl  $\beta$ -D-galactoside (300  $\mu$ g/ml). In the following, recombinant MVA viruses containing the *nef* gene and having deleted the *LacZ* marker gene were isolated by three additional consecutive rounds of plaque purification screening for non-staining viral foci in CEF cells in the presence of 5-bromo-4-chloro-3-indolyl  $\beta$ -galactoside (300  $\mu$ g/ml). Subsequently, recombinant viruses were amplified by infection of CEF monolayers, and the MVA-LAlnef viral DNA was analyzed by PCR to confirm genetic homogeneity of the virus stock. Southern blot of viral DNA confirmed genetic stability of MVA-LAlnef and precisely demonstrated integration of the *nef* gene and deletion of the *E. coli LacZ* marker gene at the site of deletion II within the viral genome. Efficient expression of recombinant Nef protein was confirmed by Western blot analysis of protein lysates from CEF cells infected with MVA-LAlnef using mouse monoclonal antibodies directed against HIV-1 Nef (kindly provided by K. Krohn and used as described by Ovod, V. et al., *AIDS*, 6:25-34 (1992)).

#### 2.4. CONSTRUCTION AND CHARACTERIZATION OF RECOMBINANT VIRUS MVA-hTYR

A 1.9 kb DNA fragment containing the entire gene encoding human tyrosinase (Tyrosinase c-DNA clone 123.B2 isolated from the melanome cell line SK29-MEL of patient

SK29 (AV), GenBank Acct. No. U01873; Brichard, V. et al.,  
*J. Exp. Med.*, 178:489-495 (1993)) was prepared from the  
plasmid pcDNAI/Amp-Tyr (Wolfel, T. et al., *Eur. J.*  
*Immunol.*, 24:759-764 (1994)) by *EcoRI* digest, modified by  
5 incubation with Klenow DNA polymerase to generate blunt  
ends, and cloned into the *SmaI* site of pUC II LZdel P7.5 to  
make the vector pUC II LZdel P7.5-TYR (Figure 6).

This plasmid could be used to engineer MVA recombinant  
virus that expresses the human tyrosinase gene under  
10 control of the vaccinia virus early/late promoter P7.5.

CEF cells infected with MVA at a multiplicity of 0.05  
TCID<sub>50</sub> per cell were transfected with DNA of plasmid pUC II  
LZdel P7.5-TYR as described previously (Sutter, G, et al.,  
*Vaccine*, 12:1032-1040 (1994)). Recombinant MVA virus  
15 stably expressing the gene for human tyrosinase and  
transiently co-expressing the *E. coli LacZ* gene was  
selected by consecutive rounds of plaque purification in  
CEF cells stained with 5-bromo-4-chloro-3-indolyl  
 $\beta$ -D-galactoside (300  $\mu$ g/ml). In the following, recombinant  
20 MVA virus expressing the gene encoding human tyrosinase and  
having deleted the *LacZ* marker gene was isolated by three  
additional consecutive rounds of plaque purification  
screening for non-staining viral foci in CEF cells in the  
presence of 5-bromo-4-chloro-3-indolyl  $\beta$ -D-galactoside (300  
25  $\mu$ g/ml). Subsequently, recombinant viruses were amplified  
by infection of CEF monolayers, and the MVA-hTYR viral DNA  
was analyzed by PCR to confirm genetic homogeneity of the

virus stock. Southern blot analysis of viral DNA confirmed genetic stability of MVA-hTYR and precisely demonstrated integration of the recombinant tyrosinase gene and deletion of the *E. coli* *LacZ* marker gene at the site of deletion II  
5 within the viral genome.

Efficient expression of recombinant human tryosinase was confirmed by Western blot analysis of protein lysates from CEF cells infected with MVA-hTYR using rabbit polyclonal antibodies (kindly provided by V. Hearing and  
10 used as described by Jimenez, M., et al., *P.N.A.S. USA*, 85:3830-3834 (1988)) or mouse monoclonal antibodies (kindly provided by L. Old and used as described by Chen, Y. et al., *P.N.A.S. USA* 92:8125-8129 (1995)) directed against tyrosinase.

## 15 EQUIVALENTS

While this invention has been particularly shown and described with references to preferred embodiments thereof, it will be understood by those skilled in the art that various changes in form and details may be made therein  
20 without departing from the spirit and scope of the invention as defined by the appended claims. Those skilled in the art will recognize or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described  
25 specifically herein. Such equivalents are intended to be encompassed in the scope of the claims.

## CLAIMS

What is claimed is:

1. A recombinant MVA virus containing and capable of  
expressing at least one foreign gene inserted at the  
5 site of a naturally occurring deletion within the MVA  
genome.
2. A recombinant MVA virus according to Claim 1  
containing and capable of expressing at least one  
foreign gene inserted at the site of deletion II  
10 within the MVA genome.
3. A recombinant MVA virus according to Claim 1 wherein  
the foreign gene codes for a marker, a therapeutic  
agent or an antigenic determinant.
4. A recombinant MVA virus according to Claim 3 wherein  
15 the foreign gene codes for an antigenic determinant  
from a pathogenic virus, a bacteria, or other  
microorganism, or from a parasite, or a tumor cell.
5. A recombinant MVA virus according to Claim 4 wherein  
the foreign gene codes for an antigenic determinant  
20 from Plasmodium Falciparum, Mycobacteria, Herpes



-39-

virus, influenza virus, hepatitis, or human immunodeficiency viruses.

6. A recombinant MVA virus according to Claims 4 wherein the antigenic determinant is HIV nef or human tryosinase.
7. A recombinant MVA virus according to Claim 6 which is MVA-LAInef or MVA-hTYR.
8. A recombinant MVA virus according to Claim 1 wherein the foreign gene codes for T7 RNA polymerase.
9. A recombinant MVA virus according to Claim 8 which is MVA-T7 pol.
10. A recombinant MVA virus according to Claim 1 wherein the foreign gene is under transcriptional control of the vaccinia virus early/late promoter P7.5.
11. Recombinant MVA viruses according to Claim 1 essentially free from viruses being able to replicate in human cells.
12. The use of a recombinant MVA virus according to Claim 8 for the transcription of DNA sequences under

-40-

transcriptional control of a T7 RNA polymerase promoter.

13. A eukaryotic cell infected by a recombinant MVA virus according to Claim 1.
- 5 14. A cell according to Claim 13 infected by a recombinant MVA virus wherein the foreign gene codes for T7 RNA polymerase.
15. A cell according to Claim 14 additionally containing one or more expression vectors carrying one or more  
10 foreign genes under transcriptional control of a T7 RNA polymerase promoter.
16. The use of cells according to Claim 15 for the production of the polypeptides encoded by said foreign genes comprising:  
15 a) culturing said cells under suitable conditions, and  
b) isolating the polypeptides encoded by said foreign genes.
17. A cell according to Claim 14 additionally containing  
20 expression vectors carrying viral genes, and/or a viral vector encoding the genome of a viral vector

-41-

under transcriptional control of a T7RNA polymerase promoter.

18. The use of cells according to Claim 17 for the production of viral particles comprising:

- 5       a)     culturing said cells under suitable conditions,  
          and  
       b)     isolating the viral particles.

19. A cell according to Claim 14 additionally containing:

- 10       a)     an expression vector carrying a retroviral vector  
          construct capable of infecting and directing the  
          expression in target cells of one or more foreign  
          genes carried by said retroviral vector  
          construct, and  
       b)     one or more expression vectors carrying the genes  
15       encoding the polypeptides required for the genome  
          of said retroviral vector construct to be  
          packaged under transcriptional control of a T7  
          RNA polymerase promoter.

20       20. The use of cells according to Claim 19 for the  
          production of retroviral particles comprising

- a)     culturing said cells under suitable conditions,  
          and  
       b)     isolating the retroviral particles.

-42-

21. A vaccine containing a recombinant MVA virus according to Claim 3 in a physiologically acceptable carrier.
22. The use of a recombinant MVA virus according to Claim 3 for the preparation of a vaccine.
- 5 23. The use of a vaccine according to Claim 21 for the immunization of a living animal body, including a human, comprising inoculation said living animal body, including a human with the vaccine.
- 10 24. The use of a vaccine according to Claim 21 containing MVA-LAlnef for the prevention or treatment of HIV infection or AIDS.
25. The use of a vaccine according to Claim 21 containing MVA-hTYR for the prevention or treatment of melanomas.
- 15 26. A vaccine comprising as a first component a recombinant MVA virus according to Claim 8 in a physiologically acceptable carrier, and as a second component a DNA sequence carrying an antigenic determinant under transcriptional control of a T7 RNA polymerase promoter in a physiologically acceptable carrier, the two components being contained together or separately.
- 20

27. The use of a vaccine according to Claim 26 for the immunization of a living animal body, including a human, comprising inoculation of said living animal body, including a human, with the first and second component of the vaccine either simultaneously or with a timelag but using the same inoculation site.
28. The use of a recombinant MVA virus according to Claim 8 for the preparation of a vaccine.
29. A recombinant MVA virus containing and capable of expressing at least one foreign gene inserted at the site of a naturally occurring deletion within the MVA genome, said recombinant MVA virus being unable to form plaques on CV1 cells.
30. A recombinant MVA virus containing and capable of expressing at least one foreign gene inserted at the site of a naturally occurring deletion within the MVA genome, said foreign gene not causing infection or disease.

-44-

RECOMBINANT MVA VIRUS, AND THE USE THEREOF

ABSTRACT OF THE DISCLOSURE

The present invention relates to recombinant vaccinia  
viruses derived from the modified vaccinia virus Ankara  
5 (MVA) and containing and capable of expressing foreign  
genes which are inserted at the site of a naturally  
occurring deletion in the MVA genome, and the use of such  
recombinant MVA viruses for the production of polypeptides,  
e.g. antigens or therapeutic agents, or viral vectors for  
10 gene therapy, and the use of such recombinant MVA viruses  
encoding antigens as vaccines.

# MVA *Hind*III

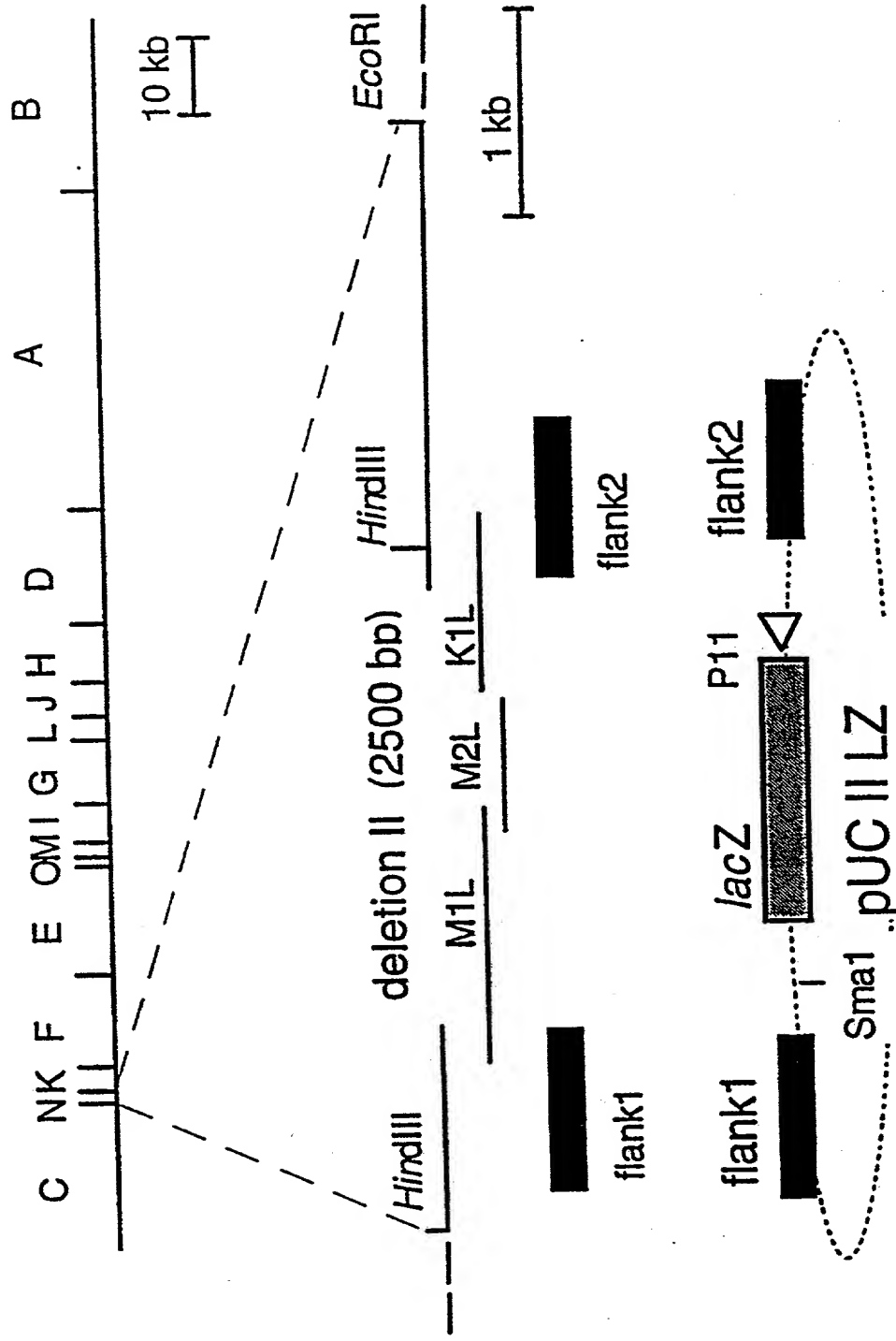


FIGURE 1

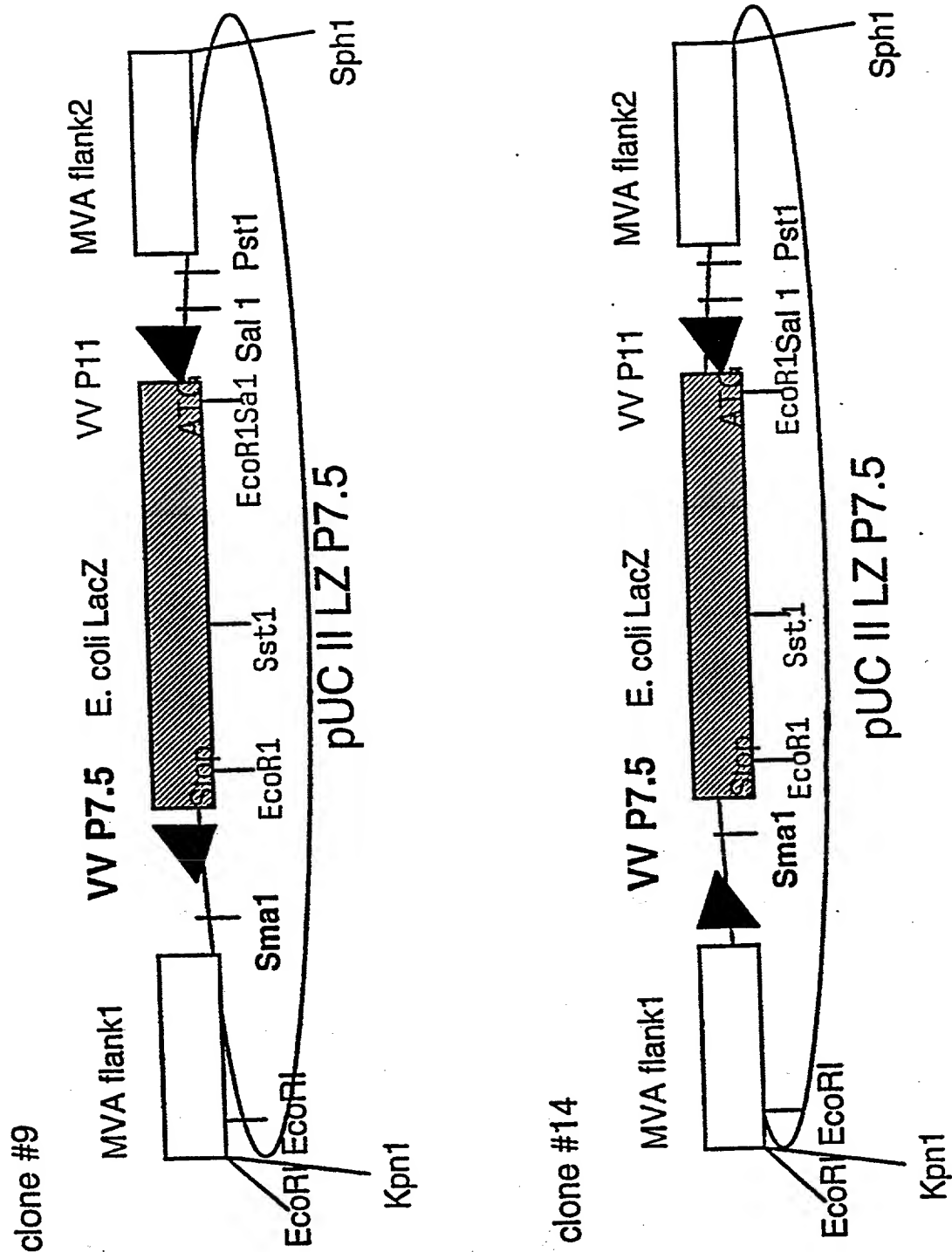


FIGURE 2



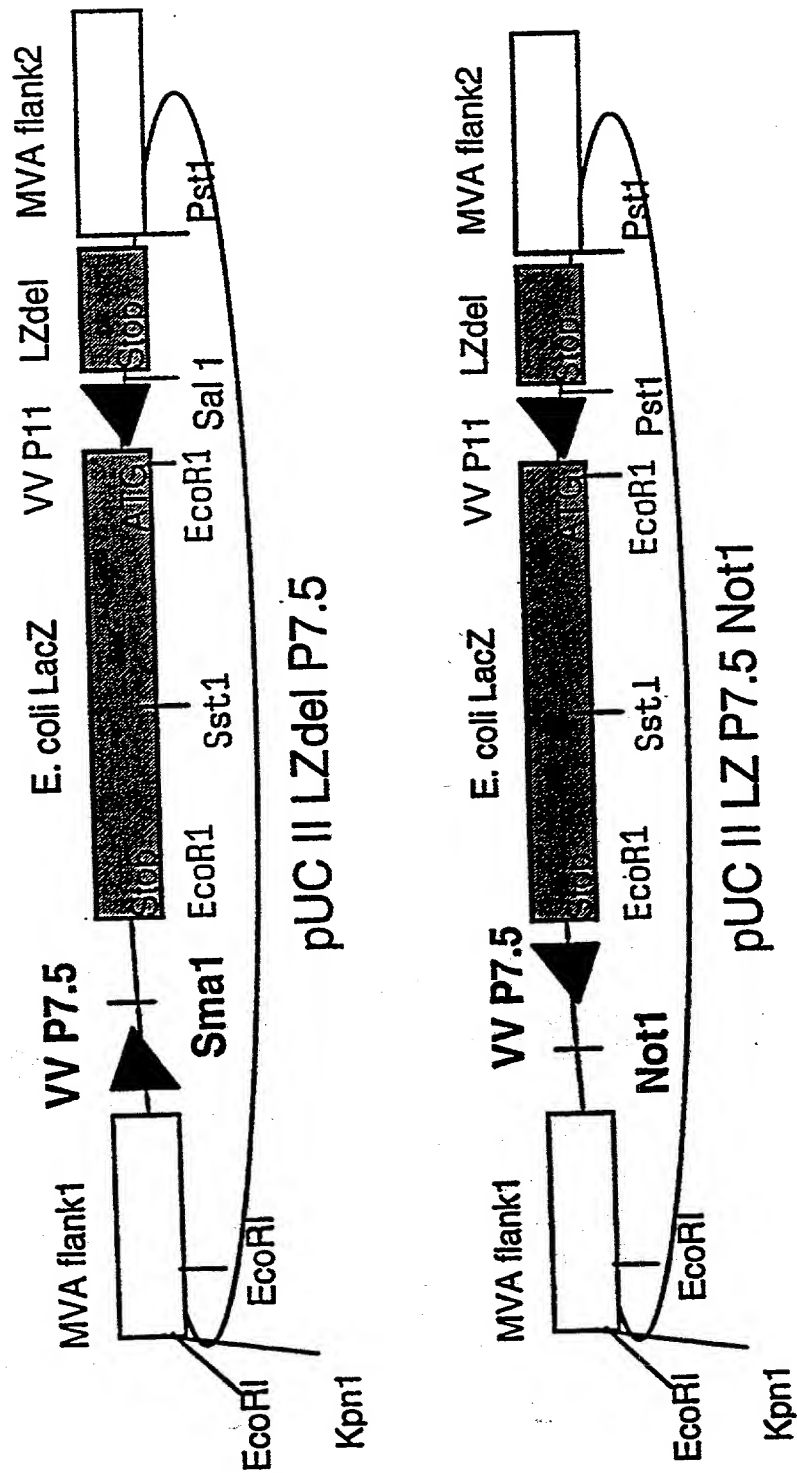


FIGURE 3

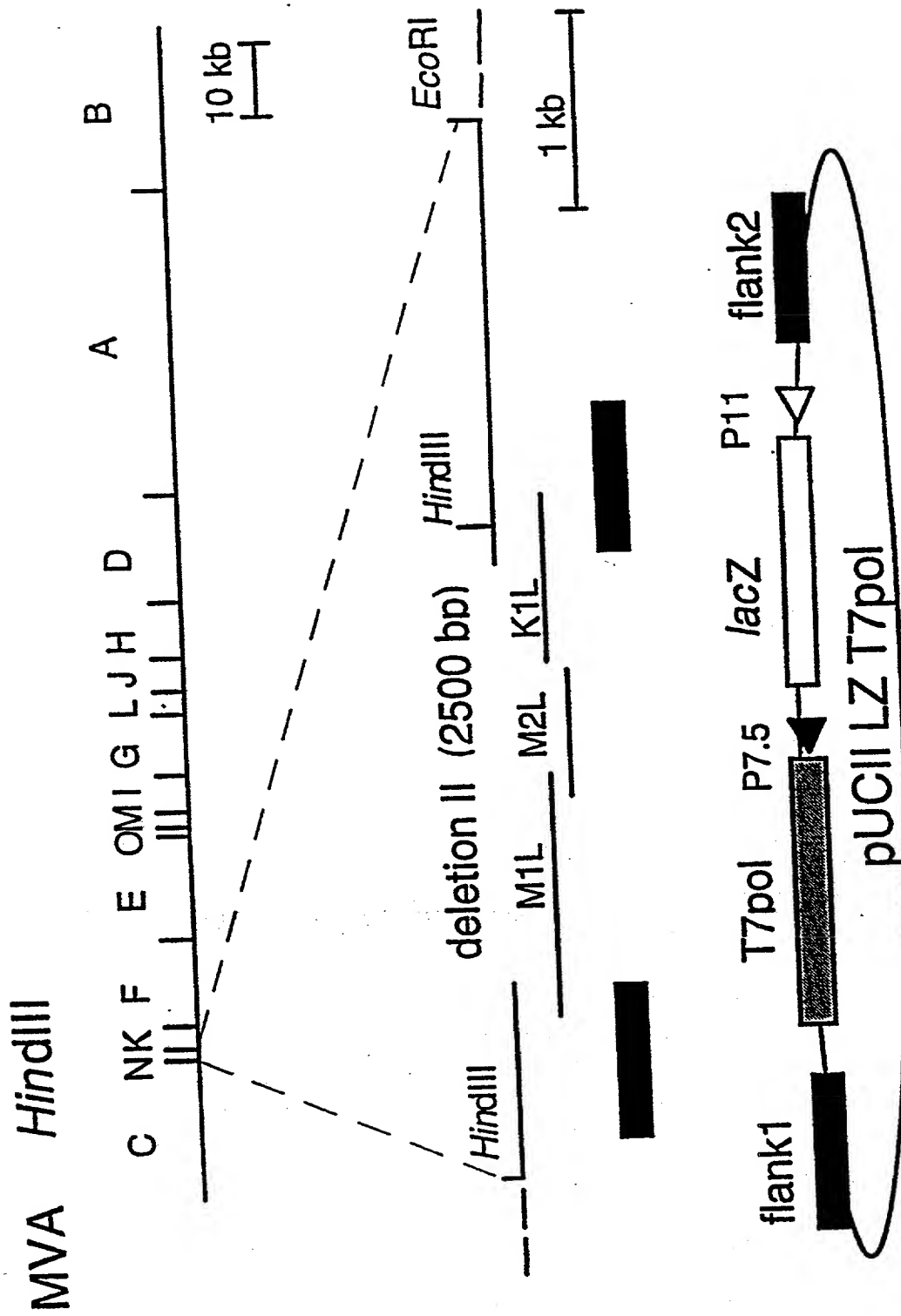


FIGURE 4

# MVA *Hind*III

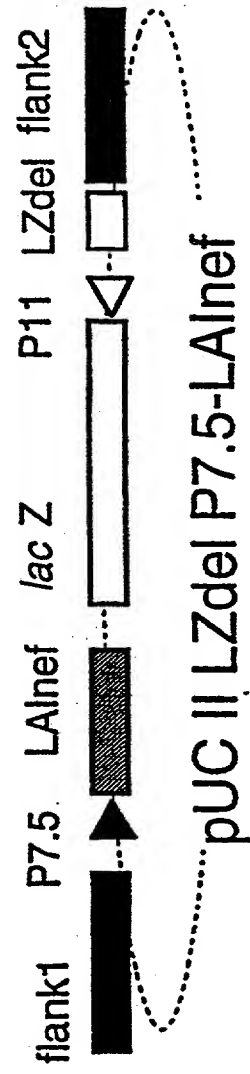
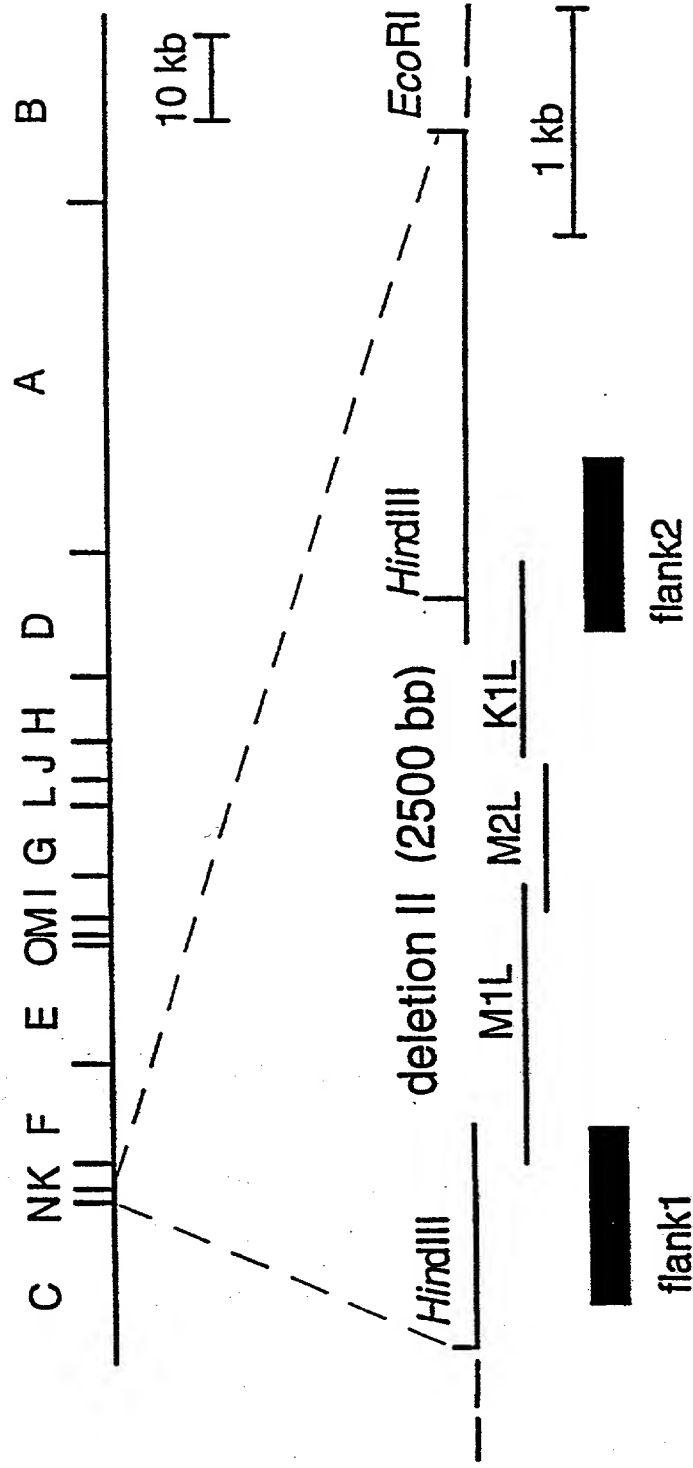


FIGURE 5

**MVA** *HindIII*

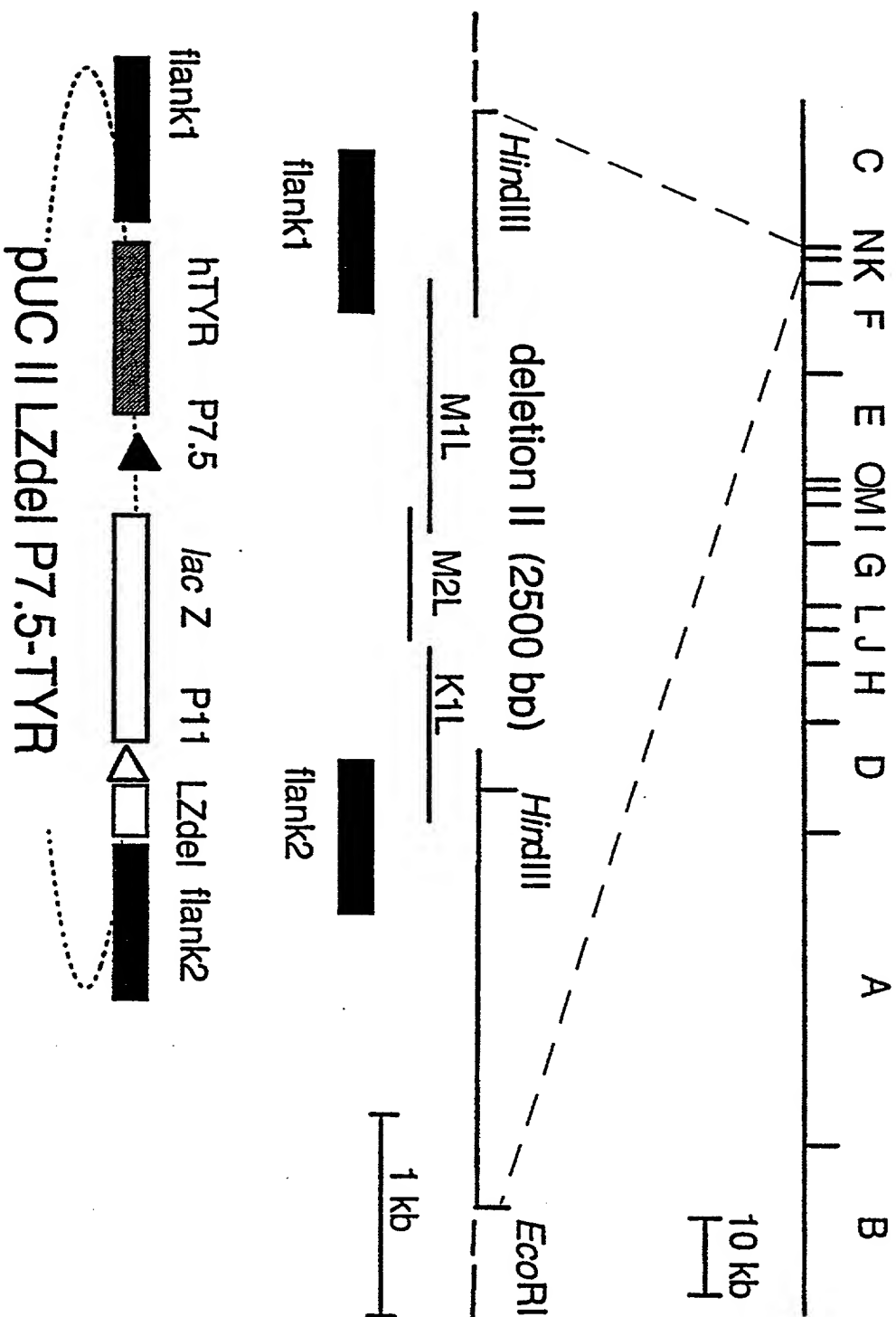


FIGURE 6